



Proximate Analysis and Phytochemical Profile of *Brachystegia eurycoma* Leaves

Uyoyoghene O. Ududua^{1*}, Michael O. Monanu¹ and Lawrence C. Chuku¹

¹Department of Biochemistry, University of Port Harcourt, Port Harcourt, Rivers State, Nigeria.

Authors' contributions

This work was carried out in collaboration among all authors. Authors UOU, MOM and LCC designed the study. Author UOU wrote the protocol, performed the statistical analysis and wrote the first draft of the manuscript. Authors MOM and LCC managed the analyses of the study. Author LCC managed the literature searches. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/AJRB/2019/v4i230064

Editor(s):

- (1) Dr. Sulaiman Shams, Assistant Professor, Department of Biochemistry, Abdul Wali Khan University Mardan, Pakistan.
(2) Dr. Khadiga Mohamed Abu-Zied, Professor, Department of Photochemistry, National Research Centre, Cairo, Egypt.

Reviewers:

- (1) Dr. Jaya Vejayan, Universiti Malaysia Pahang, Lebuhraya Tun Razak, Malaysia.
(2) Rosany Piccolotto Carvalho, Universidade Federal do Amazonas (ICB), Brazil.
Complete Peer review History: <http://www.sdiarticle3.com/review-history/48223>

Original Research Article

Received 12 January 2019

Accepted 27 March 2019

Published 03 April 2019

ABSTRACT

Aim: This study investigated the proximate and phytochemical composition of *Brachystegia eurycoma* leaves.

Methods: Crude ethanol extract of *B. eurycoma* leaves was obtained by cold extraction method. AOAC method was used for proximate analysis. Phytochemical profiling was done with qualitative phytochemical evaluation and gas chromatography-mass spectrometry (GC/MS) analytical method. Matching and interpretation of the spectral was done with the National Institute standard and Technology (NIST05) library.

Results: The proximate analysis result showed *B. eurycoma* leaves to be abundant in parameters evaluated in the order of 31.47±0.43% Carbohydrate > 15.15±0.04% Ash > 14.45±0.15 crude fibre > 13.83±0.32 protein > 13.14±0.22 moisture > 1.97±0.01 fat. Qualitative phytochemical analysis detected alkaloid, saponin, tannin, diterpenes, phenol, quinine, flavonoid, protein, xanthoprotein and cardiac glycoside in the leaves of *B. eurycoma*. GC/MS data showed that the prevailing volatile bioactive compounds in ethanol leaf extract of *B. eurycoma* were 3-O-Methyl-d-glucose (13.23%), cis-9-Hexadecenal (10.40%), Desulphosinigrin (10.34%), Phytol (7.58%), Hydroquinone (7.23%), n-Hexadecanoic acid (6.61%), Oleoyl chloride (6.10%), 9,12-Octadecadienoic acid (Z,Z)- (5.89%),

*Corresponding author: E-mail: uyoyoghene@gmail.com;

Hexadecanoic acid, (2.97%), Benzofuran, 2,3-dihydro-(1.94%), Hexadecanoic acid, 2-hydroxy-1-(hydroxymethyl) (1.92%).

Conclusion: The result of this preliminary investigation reveals the potentials of *B. eurycoma* leaves as candidate for food, pharmaceutical, cosmetic and nutraceutical industries.

Keywords: Gas chromatography mass spectrometry (GC-MS); vegetable; volatile; nutraceutical; Desulphosinigrin; drug discovery.

1. INTRODUCTION

Plants contain a wide range of bioactive chemical substances (flavonoids, alkaloids, steroids, terpenoids, phenolic acids, tannins, saponins among others) that exhibits therapeutic, physiological and biochemical effects in human body [1-4]. Scientific research resulting in phytochemical profiling, isolation, purification and characterization of phytochemicals has led to the discovery of drug candidates, production of active drugs, supplements and food additives used in the treatment and management of different ailments all over the world [5-7].

Proximate analysis is an important procedure to determine the overall composition, nutritional status, quality and energy value of any ingredient intended for use as food [8]. Preliminary phytochemical screening is used to identify the classes of bioactive constituents present in plant [9]. Gas Chromatography Mass Spectroscopy is a technique which is used to separate and identify volatile compounds based on retention time and fragmentation pattern. The fragmentation pattern for a compound is unique and is therefore an identifying characteristic of that compound [10,11]. GC-MS studies have been increasingly applied for the analysis of medicinal plants in recent year as this technique has proved to be a valuable method for the analysis of non-polar components and volatile essential oil, fatty acid, lipids and alkaloids [12].

Brachystegia eurycoma is a dicotyledonous leguminous tress belonging to the family *Fabaceae*. It is found in the swamps and rainforest of south, east and western Nigeria. It is called "Achi" by Igbos, "Ekalado" by the yorubas, "Taura" by the hausas, "Okweri" by the Edos and "Apaupan" by the Ijaw. In the eastern part of Nigeria, its seeds are used as thickener in preparation of local soups particularly egusi and ogbono soup, while its wound healing and inflammatory properties have been mentioned in literature [13].

The leaf, bark and root of the plant are used in ethno medicines for the treatment of various

diseases including malaria, diabetes, rheumatism, hypertension, kidney problem, asthma, tuberculosis, bronchitis, catarrh and sore throat [14-16].

Research has shown the seed, stem bark and stem gum as well as phytocomponents isolated from them to be anti-inflammatory, antibacterial, analgesic, antioxidant, antimicrobial, anti-cancer and anti-diarrhea [17-21]. Several researches have been done on the seeds, stem bark and stem gum of *B. eurycoma* with a little attention on the leaves which despite its medicinal use with other parts of the plant is not eaten as food. Hence this research to evaluate the nutrition and phytochemical components of *B. eurycoma* leaves with the aim of unveiling it possible nutritional and nutraceutical potentials.

2. MATERIALS AND METHODS

2.1 Plant Material

B. eurycoma leaves were collected from Abakiliki, Ebonyi State, Nigeria, authenticated by Dr. N. L. Edwin-Nwosu of the Department of Plant Science and Biotechnology, University of Port Harcourt, River State, Nigeria and assigned voucher number UPH.NO.V.1209. The leaves were rinsed in distilled water, air-dried and ground into powder.

2.2 Preparation of Ethanol Extract

The leaf powder was soaked in ethanol (70%) for 48 hours with occasional shaking. It was filtered and the filtrate concentrated after eliminating the ethanol using rotary evaporator.

2.3 Determination of Proximate Composition

The amount (%) of moisture, ash, lipid, protein, fiber, carbohydrates as well as energy level in air dried *B. eurycoma* leaf was determined in triplicate using the method of AOAC [22].

2.4 Preliminary Phytochemical Determination

The concentrated leaves extract of *B. eurycoma* were analyzed for the presence of saponins, flavonoids, tanins, alkaloids, phenols, quinines, protein, xanthoprotein, cardiac glucoside, Coumarin, steroids, diterpene and Anthraquinone using standard methods as described by Fafowora [23], Trease and Evans [24] and Harborne [25].

2.5 Gas Chromatograph-Mass Spectroscopy (GC-MS) Analysis

GC-MS was performed on a system consisting of GC2010 gas chromatograph and a Shimadzu QP2010 ultra quadrupole mass spectrometer equipped with a DB-Wax fused silica capillary column (30m x 0.25mm ID x 0.25µm df, composed of silphenylene polymer). Initial temperature was programmed at 50°C, held for two minutes. It was increased to 300°C with the rate of 6.5°C/min and held for ten minutes. The temperature of the injector and detector were set up to 280°C and 300°C, respectively. Helium gas was used as a carrier gas. 1 µl of the fractions was diluted in 200 µl dichloromethane and then injected into the GC-MS [26,27].

2.6 Identification of Spectral

Interpretation of mass-spectrum was done by comparing the mass spectrum of the unknown component with spectrum of known components in the National Institute Standard and

Technology (NIST05) database to ascertain the name, molecular weight and structure of the components of the test materials.

3. RESULTS

The result of proximate analysis shows that the leaves of *B. eurycoma* to contain 31.47±0.43% carbohydrates, 15.15±0.04% ash, 14.45±0.15% crude fiber, 13.83±0.32% protein, 13.14±0.22% moisture, 1.97±0.01% fat and 198.87±0.56 Kcal/100g (Table 1).

Table 1. Proximate composition of *B. eurycoma* leaves

Parameter	Unit	<i>B. eurycoma</i>
Carbohydrate (NFE)	%	31.47±0.43
Ash	%	15.15±0.04
Crude fibre	%	14.45±0.15
Protein	%	13.83±0.32
Moisture	%	13.14±0.22
Fat	%	1.97±0.01
Energy level	Kcal/100g	198.87±0.56

Qualitative phytochemical analysis revealed the presence of alkaloid, saponin, tannin, diterpenes, phenol, quinine, flavonoid, protein, xanthoprotein and cardiac glycoside in the leaves of *B. eurycoma*. Coumarin, steroids and anthraquinone were not detected (Table 2).

The GC-MS chromatogram obtained from ethanol extract of *B. eurycoma* shows the presence of 24 distinct peaks (Fig. 1).

Table 2. Qualitative phytochemical evaluation of *B. eurycoma* leaves

	Phytochemical components	Type of test	Ethanol leaf extract of <i>B. eurycoma</i>
1	Alkaloids	Mayer's Test Wanger's test Dragendroff's test	+ + +
2	Phenol	Ferric Chloride	+
3	saponin	Frothing	+
4	Tannin	Ferric Chloride	+
5	Coumarin	alcoholic sodium hydroxide	-
6	Quinine	potassium hydroxide	+
7	Anthraquinone	Borntrager's	-
8	steroids	Liebermann- Burchard	-
9	Protein	Million's	+
10	Xanthoprotein	General test	+
11	Cardiac glycoside	Keller kiliani	+
12	Flavonoid	sodium hydroxide	+
13	Diterpene	Copper acetate ²⁵	+

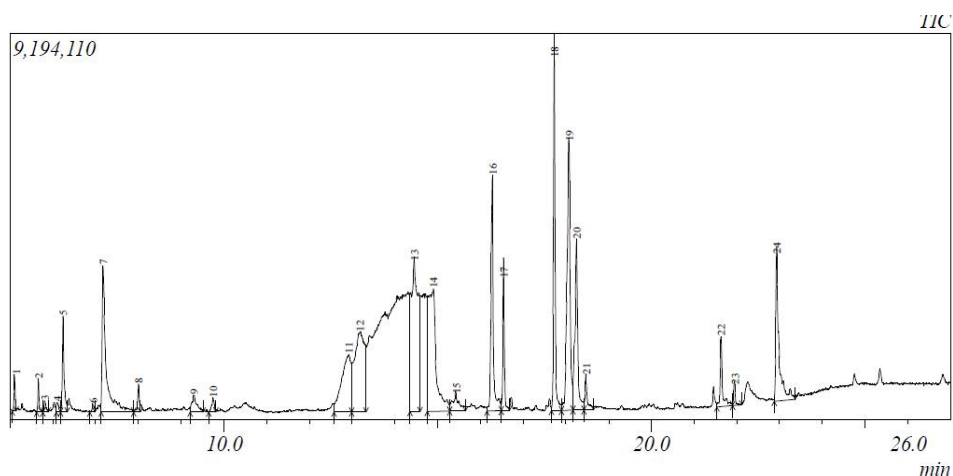

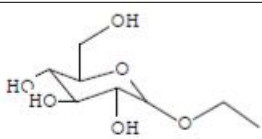
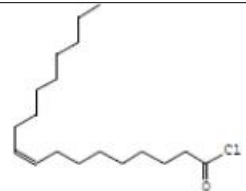
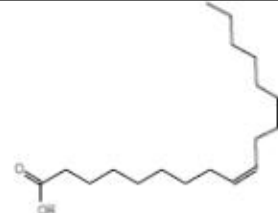
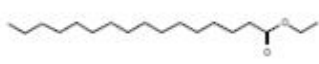
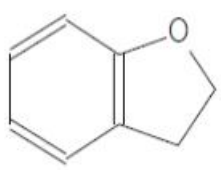
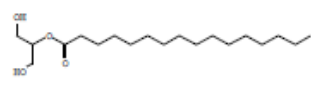
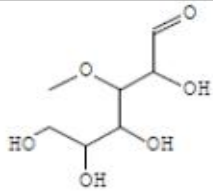
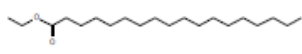
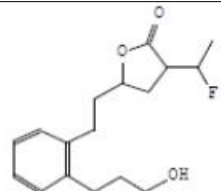
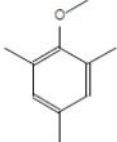
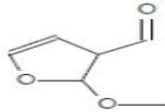
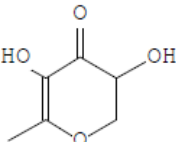
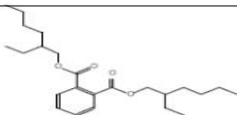
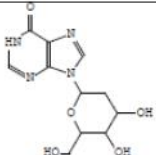
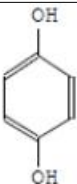
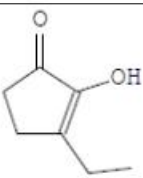
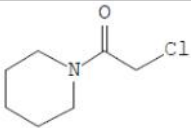


Fig. 1. GC-MS chromatogram of ethanol leaf extract of *B. eurycoma*

Table 3. Molecular formula and structure of phytochemicals identified in *B. eurycoma*

Name of compound	Molecular formula	Structure	Molecular weight	Peak Area (%)
1 3-O-Methyl-d-glucose	C ₇ H ₁₄ O ₆		194	13.23
2 3-O-Methyl-d-glucose	C ₇ H ₁₄ O ₆		194	12.42
3 cis-9-Hexadecenal ; 9-Hexadecenal, (Z)-	C ₁₆ H ₃₀ O		238	10.40
4 Desulphosinigrin	C ₁₀ H ₁₇ NO ₆ S		279	10.34
5 Phytol	C ₂₀ H ₄₀ O		296	7.58
6 Hydroquinone	C ₆ H ₆ O ₂		110	7.23

7	n-Hexadecanoic acid	$C_{16}H_{32}O_2$		256	6.61
8	Ethyl .alpha.-d-glucopyranoside	$C_8H_{16}O_6$		208	6.61
9	Oleoyl chloride	$C_{18}H_{33}ClO$		300	6.10
10	9,12-Octadecadienoic acid (Z,Z)-	$C_{18}H_{32}O_2$		280	5.89
11	Hexadecanoic acid, ethyl ester ;	$C_{18}H_{36}O_2$		284	2.97
12	2,3-dihydro-Benzofuran Coumaran	C_8H_8O		120	1.94
13	Hexadecanoic acid, 2-hydroxy-1-(hydroxymethyl)ethyl ester	$C_{19}H_{38}O_4$		330	1.92
14	Palmitin, 2-mono-3-O-Methyl-d-glucose	$C_7H_{14}O_6$		194	1.43
15	Octadecanoic acid, ethyl ester	$C_{20}H_{40}O_2$		312	1.05
16	Tetrahydrofuran-2-one, 3-[1-fluoroethyl]-5-[[2-hydroxypropyl]benzen eethyl-	$C_{17}H_{23}FO_3$		294	0.99

17	Benzene, 2-methoxy-1,3,5-trimethyl-	$C_{10}H_{14}O$		150	0.56
18	Furan-3-carboxaldehyde 2-methoxy-2,3-dihydro-	$C_6H_8O_3$		128	0.50
19	4H-Pyran-4-one 2,3-dihydro-3,5-dihydroxy-6-methyl-	$C_6H_8O_4$		144	0.50
20	Bis(2-ethylhexyl) phthalate	$C_{24}H_{38}O_4$		390	0.49
21	9-[2-Deoxy-.beta.-d-ribohexopyranosyl]purin-6(1H)-one	$C_{11}H_{14}N_4O_5$		282	0.45
22	Hydroquinone	$C_6H_6O_2$		110	0.31
23	2-Cyclopenten-1-one 3-ethyl-2-hydroxy-	$C_7H_{10}O_2$		126	0.26
24	Piperidine, 1-chloroacetyl-	$C_7H_{12}ClNO$		161	0.20

The phytochemicals identified in order of most abundant to least abundant based on percentage peak are presented in Table 3, showing their molecular formula and structure.

4. DISCUSSION

The findings from proximate analysis of this study shows *B. eurycoma* leaves to have the abundance of the parameters evaluated to be in Carbohydrate > Ash > crude fiber > protein > moisture > fat order. This shows *B. eurycoma* leaves are a better source of carbohydrates and

fiber than proteins. Animals depend on carbohydrates among other macromolecules for generation of energy and some intermediates required for certain biological processes and the sustenance of life. The amount of carbohydrates (31.47%) is appreciable and similar to popular edible vegetables like *Celusia argenta* (32.80%) and *Corchorus olitorius* (31.30%) [28]. The ash content of the leaves (15.15%) is considerable showing that the leaves contain important mineral elements as the ash content of any sample is an index of mineral content.

The Recommended Dietary Allowance (RDA) of fiber is 19-25% for children, 21-38% for adult, 28% for pregnant mothers and 29% for breast-feeding mothers [29]. With crude fibre of 14.45%, *B. eurycoma* leaves will make a poor source of dietary fiber in human nutrition. Plant foods that provide more than 12% of its calorific value from protein is considered good source of protein [30]. *B. eurycoma* leaves meet this requirement, making it a good source of protein. Moisture content was lower when compared with that of common vegetable such as *Telfairia occidentalis* 98%, *Talinum triangulare* 91%, *Moringa oleifera* 87% and *Vernonia amygdalinan* 87% [31] but low moisture content is indicative of a longer shelf life. It is a general observation that leafy vegetables have low lipid content [30]. The low lipid content of *B. eurycoma* leaves is in agreement with this observation.

Preliminary phytochemical screening detected the presence of alkaloid, saponin, tannin, diterpenes, phenol, quinine, flavonoid, protein, xanthoprotein and cardiac glycoside. Alkaloids are diuretic in nature, they affect the nervous system and reduce appetite [32]. The ingestion of saponins as a part of the human diet have been linked with a variety of effects on health, including reducing blood cholesterol levels. They have also been reported to have pharmacological activities such as anti-inflammatory, antifungal, antibacterial, anti-parasitic, anti-cancer and antiviral activities [33]. Tannins are antiinflammatory, antidiarrheal, haemostatic, antiviral, antibacterial, antidiarrheal and antihemorrhoidal compounds which has been reported to relief sore throat, fatigue and skin ulcer [34]. Diterpenes are antimicrobial and anti-inflammatory [35]. Quinine has many medicinal applications due to its fever-reducing, painkilling and anti-inflammatory properties [36]. Many flavonoids have been shown to have antioxidative activity, free radical scavenging capacity, antihypertensive, hepatoprotective, anti-inflammatory, antiviral and anticancer activities [37]. Cardiac glycosides have cardiotoxic activity, are antiviral, anticancer and antiproliferative effects [38].

GC-MS analysis of ethanol leaf extract of *B. eurycoma* revealed the presence of 24 phytochemical compounds; Amongst which were the sugar moiety (3-O-Methyl-d-glucose and Ethyl .alpha.-d-glucopyranoside), aldehyde (cis-9-Hexadecenal, Furan-3-carboxaldehyde-2-methoxy-2,3-dihydro-), Glucosinolates (Desulphosinigrin), Diterpene (phytol), phenol

(Hydroquinone, 2,3-dihydro- Benzofuran), fatty acids esters (n-Hexadecanoic acid, 9,12-Octadecadienoic acid (Z,Z)-, Hexadecanoic acid, Hexadecanoic acid, 2-hydroxy-1-(hydroxymethyl), Octadecanoic acid) and Flavonoid (4H-Pyran-4-one-2,3-dihydro-3,5-dihydroxy-6-methyl-). The detection of sugar moieties by GC-MS analysis is in agreement with the high amount of carbohydrate observed in proximate analysis.

Phytochemical and ethnobotanical database has ascribed several pharmacological or biological activities such as anti-inflammatory, antioxidant, antilipidemic, antihistaminic, antimicrobial amongst others to majority of the compounds identified [39]. 3-O-Methyl-d-glucose, the most abundant component in *B. eurycoma*, is a non-metabolizable glucose analog, used as a proxy for d-glucose uptake in *in vivo* absorption studies and as a preservative [40,41].

Research findings have reported the anticancer and antimicrobial nature of desulphosinigrin [42]; antioxidant and antifungal activities of 4H-Pyran-4-one,2,3-,dihydro-3,5-dihydroxy-6-methyl-[43-45]; 5-Alpha reductase inhibitor, anti-androgenic, anticancer, antioxidant, hypocholesterolemic, acne reductive, anti-inflammatory and anti-eczemic effect of 9,12-Octadecadienoic acid (Z,Z)- [46-48], as well as the anti-inflammatory, anti-cancer, cytotoxic and antimicrobial properties of n-Hexadecanoic acid and Hexadecanoic acid [49-51].

Phytol, a building block of chlorophyll, is among the twenty four compounds identified in the present study. It is used in the manufacture of synthetic vitamins E and K₁. It has been reported to have antinociceptive, antioxidant, anti-inflammatory, antiallergic, immunostimulant, antimicrobial, antischistosoma, cancer preventive, sedative and anxiolytic effects [52-60]. Hydroquinone occurs naturally in various medicinal plants [61,62], it has been reported to be allelochemic, antimicrobial, antihepatic, antilithic, antimelanomic, antimelasmaic, uroantiseptic, antimitotic, and antipertussive [63-65]. Benzofuran, 2,3-dihydro is a coumaran and research has shown that it possesses anti-inflammatory, antidiarrheal, antileishmanial, immunomodulatory, antimicrobial and anti-helminthic activities [66-68]. Octadecanoic acid is a flavouring agent which is hypocholesterolemic [69]. 2,3-dihydro-3,5-dihydroxy-6-methyl-4H-Pyran-4-one, a flavonoid with antifungal, antioxidant [70-71].

5. CONCLUSION

It could be seen, based on constituents detected in this preliminary study, that *B. eurycoma* leaves are nutritious and contains volatile phytochemicals with known biological activities revealing it as a potential candidate in food, pharmaceutical, cosmetic and nutraceutical industries. This research serves as bases for more research on *B. eurycoma* leaves.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Okwu DE, Omodamiro OD. Effects of hexane extract and phytochemical content of *Xylopiya aethiopica* and *Ocimum gratissimum* on the uterus of guinea pig. *Bio Res* 2005;3(2):40-4.
- Yousuf S, Bachheti RK, Joshi A, Mathur A. Evaluation of antioxidant potential and phytochemicals of *Morina longifolia*. *Int J Pharm Pharm Sci.* 2014;6(6):208-12.
- Zeng CC, Liu X, Chen GR, Wu QJ, Liu WW, Luo HY, Cheng JG. The molecular mechanism of Rhein in diabetic nephropathy. *Evidence-Based Complementary and Alternative Medicine.* 2014;Volume 2014, Article ID 487097, 6 pages.
- Nyamai DW, Arika W, Ogola PE, Njagi ENN, Ngugi MP. Medicinally important phytochemicals: An updated research avenue. *J. Pharmacogn. Phytochem.* 2016;4(1):35-49.
- Li J, Vederas JC. Drug discovery and natural products: End of an era or an endless frontier? *Science.* 2009;325:161-165.
- Newman DJ, Cragg GM. Natural products as sources of new drugs over the 30 years from 1981 to 2010. *J. Nat. Prod.* 2012;75: 311-335.
- Newman DJ, Cragg GM. Natural products as sources of new drugs over the last 25 years *J. Nat. Prod.* 2007;70(2007):461-477.
- Qayyum MMN, Butt MS, Anjum FM, Nawaz H. Composition analysis of some selected legumes for protein isolates recovery. *J. Anim. Plant Sci.* 2012;22: 1156-1162.
- Yadav MA, Chatterji S, Gupta SK, Watal G. Preliminary phytochemical screening of six medicinal plants used in traditional medicine. *Int J Pharm Pharm Sci.* 2014; 6(5): 539-542.
- Rajeswari J, Rani S. GC-MS Analysis of Phytochemical Compounds in the Ethanolic Extract of Root of *Lawsonia inermis* Linn. *Int.J. ChemTech Res.* 2015; 07(01);389-399.
- Mythili K, Umamaheswara RC, Chamundeeswari D, Manna PK. GC-MS analysis of phytocomponents and in vitro inhibitory effects of *Calanthe triplicata*. *J Nat Prod.* 2013; 6: 141-146.
- Kavitha S, Lincy PM, Kala MJS, Mohan VR. GC-MS analysis of ethanolic extract of *Nothapodytes nimmoniana* (Graham) Mabb leaves. *Malaya J Biosci.* 2014; 2(1):42-49.
- Uhegbu FO, Onwuchekwa CC, Iweala EJ, Kanu I. Effects of processing methods on nutritive and anti-nutritive properties of *Brachystegia eurycoma* and *Detarium microcarpum* from Nigeria. *Pakistan J Nutr.* 2009; 8: 316-20.
- Adikwu MU, Enabeke TC. Ethnomedicinal uses of *Brachystegia eurycoma*. *Animal Research International.* 2007; 4:685-697.
- Burkill HM. *The Useful Plants of West Tropical Africa.* Royal Botanical Garden, Kew. 1985; 3:120.
- Idu M, Onyibe HI. Medicinal Plants of Edo State, Nigeria. *Research Journal of Medicinal Plant.* 2007; 1(2): 32-41.
- Kunle, OO. The Production of Pharmaceuticals from Medicinal Plants and their Product. *Nat. Prod. and Medicines.* 2000; 4:9-12.
- Igbe I, Ayinde BA, Izuchukwu A. Anti-inflammatory and Analgesic Effects of the Methanol Stem Bark Extract of *Brachystegia eurycoma* Harms (Fabaceae). *Euro J Med Plants.* 2012; 2(4): 356-365.
- Igwe OU, Okwu DE. (2013). GC-MS Evaluation of bioactive compounds and antibacterial activity of the oil fraction from the stem bark of *Brachystegia eurycoma* Harms. *Int. J. Chem. Sci.* 2013; 11(1):357-371.
- Igwe OU, Okwu DE. Isolation, characterization and antibacterial activity of 3-hydroxy-2,2-bis (6-methoxy-3-methyl-2,3-dihydrobenzofuran-2-yl) propanal from the stem exudate of *Brachystegia*

- eurycoma* Harms. Der Pharma Chemica. 2013;5(2):39-44.
21. Igbe I, Inarumen GO. The effect of leaf aqueous extract of *Brachystegia eurycoma* Harms (Fabaceae) in acute and chronic inflammatory animal models. British J Pharm Res. 2013;3(3):391-400.
 22. AOAC. Official methods of proximate analysis, AOAC International, Gaithersburg, MD. 2010;15.
 23. Sofowora A. Medicinal Plants and Traditional Medicine in Africa. 2nd ed. Ibadan, Nigerian: Spectrum Books Ltd.; 1993;289.
 24. Trease GE, Evans WC. Pharmacognosy: a Physician's Guide to Herbal Medicine. 13th ed. London: Bailliere Tindall. 1989;176-80.
 25. Harborne JB. Phytochemical methods. A Guide to modern technique of plant analysis. London: Chapman and Hall Ltd.; 1973;49-188.
 26. Houghton PJ, Raman A. Laboratory handbook for the fractionation of natural extracts. 1st ed. London: Chapman and Hall; 1998.
 27. Pavia DL, Lampman GM, Kriz GS, Engel RG. Introduction to organic laboratory techniques: A microscale approach. 4th Ed. St. Paul: Cengage Learning, Brooks/Cole Publishing Co; 2006.
 28. Onwordi CT, Ogungbade AM, Wusu AD. The proximate and mineral composition of three leafy vegetables commonly consumed in Lagos, Nigeria. Afr J Pure and Appl Chem. 2009;3(6):102-107.
 29. Idris S. Compositional Studies of *Telfairia occidentalis* Leaves. Am J Chem. 2011;1(2):56-59.
 30. Okewole SA, Oyekunle LO, Akande OO, Adebisi TT, Olubode TP. Nutritional compositions of selected green leafy vegetables in Oyo State, Nigeria. Asian J Appl. Chem. Res. 2018;1(1):1-7.
 31. Saidu AN, Jideobi NG. The proximate and elemental analysis of some leafy vegetables grown in Minna and environs. J. Appl. Sci. Environ. Manage. 2009;13(4): 21-22.
 32. United States Department of Agriculture. Center for Nutrition Policy and Promotion. Dietary Guidelines for Americans. National Academy Press, Washington DC: USA; 2010.
 33. Podolak I, Galanty A, Sobolewska D. Saponins as cytotoxic agents: A review. Phytochem Rev. 2010;6:425-474.
 34. Ashok PK, Upadhyaya K. Tannins are astringent. J Pharmacognosy and Phytochemistry. 2012;1(3):45-50.
 35. Roncero AM, Tobal IE, Moro RF, Díeza D, Marcos IS. Halimane diterpenoids: Sources, structures, nomenclature and biological activities. Nat. Prod. Rep. 2018; 35:955-991.
 36. Dawidowicz AL, Bernacik K, Typek R, Stankevič M. Possibility of quinine transformation in food products: LC-MS and NMR techniques in analysis of quinine derivatives. Eur Food Res Technol. 2018; 244(1):105-116.
 37. Kumar S, Pandey AK. Chemistry and biological activities of flavonoids: An overview. The Scientific World Journal. 2013;2013(162750).
 38. Schneider NFZ, Cerella C, Simões CMO, Diederich M. Anticancer and Immunogenic Properties of Cardiac Glycosides Molecules. 2017;22(11):1932-40.
 39. Duke JA. Handbook of phytochemical constituents of GRAS herbs and other economic plants. Boca Raton, FL. CRC Press; 1992.
 40. Liu J, Lee GY, Lawitts JA, Toner M, Biggers JD. Preservation of mouse sperm by convective drying and storing in 3-O-methyl-D-glucose. PloS One. 2012;7(1): e29924
 41. McWhorter TJ, Green AK, Karasov WH. Assessment of radiolabeled D-glucose and the nonmetabolizable analog 3-O-methyl-D-glucose as tools for in vivo absorption studies. PBZ. 2010;83(2):376-384.
 42. Krishnaveni M. Docking, simulation studies of desulphosinigrin – Cyclin Dependent Kinase 2, an Anticancer Drug Target. Int. J. Pharm. Sci. Rev. Res. 2015; 30(2):115-118.
 43. Čechovská L, Cejpek K, Konečný M, Velíšek J. On the role of 2,3-dihydro-3,5-dihydroxy-6-methyl-(4H)-pyran-4-one in antioxidant capacity of prunes. Eur Food Res Technol. 2011;233(3):367-376.
 44. Yu X, Zhaob M, Shitong FL, Hu ZJ. Identification of 2,3-dihydro-3,5-dihydroxy-6-methyl-4H-pyran-4-one as a strong antioxidant in glucose-histidine Maillard reaction products. Food Res Intern. 2013;51(1):397-403.
 45. Teoh YP, Don MM. Mycelia growth and production of total flavonoids and 4H-pyran-4-one, 2,3-dihydro-3,5-dihydroxy-6-methyl- by *Schizophyllum commune* using a bubble column bioreactor considering

- aeration effect and mass transfer study. Chem. Biochem. Eng. Q. 2014;28(4):553–559.
46. Liang T, Liao S. Inhibition of steroid 5 β -reductase by specific aliphatic unsaturated fatty acids. The Biochemical journal. 1992; 285(2):557-62.
 47. Elsayed HH, Abd Elrahman MK, Emara AH, El-Hafez A. Compare effect of fatty acid composition (Olive, Coconut Oil and Butter) on adipose liver tissue, and serum lipid profile in albino rats. J Biotech Biochem. 2015;1(3):28-38.
 48. Oloyede GK. Antioxidant activities of methyl ricinoleate and ricinoleic acid dominated *Ricinus communis* seeds extract using lipid peroxidation and free radical scavenging methods. Res J Med Plants. 2012;6:511-520.
 49. Aparna V, Mandal P, Sadasivan C, Vijayan D, Karthe P, Haridas M. Anti-inflammatory property of n-hexadecanoic acid: Structural evidence and kinetic assessment. Chem Biol Drug Design. 2012;80(3):434-9.
 50. Ravi L, Krishnan K. Cytotoxic potential of N-hexadecanoic acid extracted from *Kigelia pinnata* Leaves. Asian J Cell Biol. 2017;12:20-27.
 51. Cartron ML, England SR, Chiriac AL, Josten M, Turner R, Rauter Y, Hurd A, Sahl H, Jones S, Foster SJ. Bactericidal activity of the human skin fatty acid cis-6-hexadecanoic acid on *Staphylococcus aureus*. Antimicrobial Agents and Chemotherapy. 2014;58(7):3599–3609.
 52. Santos CCMP, Salvadori MS, Mota VG, Costa LM, Almeida AAC, Oliveira GAL, Costa JP, Sousa DP, Freitas RM, Almeida RN. Antinociceptive and antioxidant activities of phytol *in vivo* and *in vitro* models. Neurosci J; 2013. DOI:doi.org/10.1155/2013/949452.
 53. Ryu KR, Choi JY, Chung S, Kim DH. Anti-scratching behavioral effect of the essential oil and phytol isolated from *Artemisia princeps* Pamp. in mice. Planta Med. 2011;77:22–26.
 54. 54] Lim SY, Meyer M, Kjonaas RA, Ghosh SK. Phytol-based novel adjuvants in vaccine formulation: 1. assessment of safety and efficacy during stimulation of humoral and cell-mediated immune responses. J Immune Based Ther Vaccines; 2006. DOI:10.1186/1476-8518-4-6.
 55. Chowdhury RR, Ghosh SK. Phytol-derived novel isoprenoid immunostimulants. Front Immunol; 2012. DOI:10.3389/fimmu.2012.00049.
 56. Rajab MS, Cantrell CL, Franzblau SG, Fischer NH. Antimycobacterial activity of (E)-phytol and derivatives: A preliminary structure-activity study. Planta Med. 1998; 64:2–4.
 57. Saikia D, Parihar S, Chanda D, Ojha S, Kumar JK, Chanotiya CS, Shanker K, Negi AS. Antitubercular potential of some semisynthetic analogues of phytol. Bioorg Med Chem Lett. 2010;20:508–512
 58. Inoue Y, Hada T, Shiraiishi A, Hirose K, Hamashima H, Kobayashi S. Biphasic effects of geranylgeraniol, teprenone, and phytol on the growth of *Staphylococcus aureus*. Antimicrob Agents Chemother. 2005;49(5):1770–4
 59. De Moraes J, De Oliveira RN, Costa JP, Junior AL, De Sousa DP, Freitas RM, Allegretti SM, Pinto P L. Phytol, a diterpene alcohol from chlorophyll, as a drug against neglected tropical disease Schistosomiasis Mansonii. PLoS Negl Trop Dis; 2014. DOI:10.1371/journal.pntd.0002617.
 60. Costa JP, De Oliveira GA, De Almeida AA, Torequillslam M, Sousa DP, Freitas RM. (2014). Anxiolytic-like effects of phytol: Possible involvement of GABAergic transmission. Brain Res. 2014;1547:34-42.
 61. De Arriba SG1, Naser B, Nolte KU. Risk assessment of free hydroquinone derived from *Arctostaphylos Uva-ursi* folium herbal preparations. Int J Toxicol. 2013;32(6):442-53.
 62. Jurica K, Karačonji IB, Šegan S, Milojković OD, Kremer D. Quantitative analysis of arbutin and hydroquinone in strawberry tree leaves by GC-MS. Arh Hig Rada Toksikol. 2015;66:197-202.
 63. King AG, Landreth KS, Wierda D. Bone marrow stromal cell regulation of B-lymphopoiesis. II. Mechanisms of hydroquinone inhibition of pre-B cell maturation. J Pharmacol Exp Ther. 1989; 250(2):582-590.
 64. Strapkova A, Jahodar L, Nosal'ova G. Antitussive effect of arbutin. Pharmazie. 1991;46(8):611-612.
 65. Maeda K, Fukuda M. Arbutin: mechanism of its depigmenting action in human melanocyte culture. J Pharmacol Exp Ther. 1996;276(2):765-769.

66. De Castro Oliveira LG, Brito LM, De Moraes Alves MM, Amorim LV, Sobrinho-Júnior EP, De Carvalho CE, Da Franca Rodrigues KA, Arcanjo DD, Das Graças Lopes Citó AM, De Amorim Carvalho FA. *In vitro* effects of the neolignan 2,3-dihydrobenzofuran against leishmania amazonensis. Basic Clin Pharmacol Toxicol. 2017;120(1):52-58.
67. Sánchez-Ramos M, Bahena SM, Romero-Estrada A, Bernabé-Antonio A, Cruz-Sosa F, González-Christen J, Acevedo-Fernández JJ, Perea-Arango I, Alvarez L. Establishment and phytochemical analysis of a callus culture from *Ageratina pichinchensis* (Asteraceae) and its anti-inflammatory activity. Molecules. 2018; 25:23(6). DOI:10.1371/journal.pntd.0002617
68. Ravi K, Selvam K, Swaminathan M. Photochemical synthesis and antimicrobial screening of some substituted dihydrobenzofurans. M. Res Chem Intermed. 2012;38(9):2393–2400.
69. Mensink RP. Effects of stearic acid on plasma lipid and lipoproteins in humans. Lipids. 2006;40(12):1201-5.
70. Teoh Y P, Mat Don M. Mycelia Growth and Production of Total Flavonoids and 4H-pyran-4-one, 2,3-dihydro-3,5-dihydroxy-6-methyl- by *Schizophyllum commune* Using a Bubble Column Bioreactor Considering Aeration Effect and Mass Transfer Study. Chem. Biochem. Eng. Q. 2014;28(4): 553–559.
71. Yu X, Zhao M, Liu F, Zeng S, Hu J. Identification of 2,3-dihydro-3,5-dihydroxy-6-methyl-4H-pyran-4-one as a strong antioxidant in glucose–histidine Maillard reaction products. Food Res Inter. 2013;51:397–403.

© 2019 Ududua et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
The peer review history for this paper can be accessed here:
<http://www.sdiarticle3.com/review-history/48223>