



Hermaphroditism in Kiwifruit (*Actinidia deliciosa*): A Review

Shashi K. Sharma ^{a*}

^a Department of Fruit Science, College of Horticulture and Forestry, Dr. Y. S. Parmar University of Horticulture and Forestry, Neri, Hamirpur, Himachal Pradesh, 177001, India.

Author's contribution

The sole author designed, analysed, interpreted and prepared the manuscript.

Article Information

DOI: 10.9734/IJPSS/2024/v36i54525

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/114445>

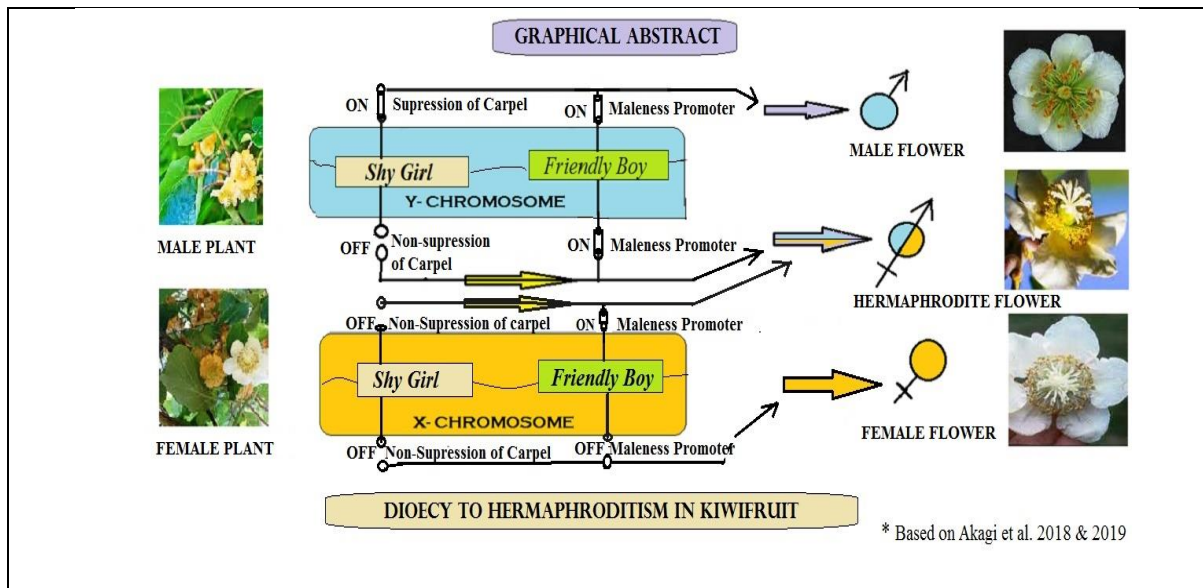
Review Article

Received: 16/01/2024
Accepted: 21/03/2024
Published: 26/03/2024

ABSTRACT

Actinidia, the kiwifruit is approximately 20 to 26 million years old. It is known widely for its dioecious nature; hermaphroditism has also been found to exist in its male inconstant type. Floral development in this plant species progresses through seven unique stages, following the conventional (A) BCE floral model. Male parts of the flower develop early during the third stage, whereas, the degeneration of these parts (pollens) in female plants occurs during later stages owing to programmed cell death. After exploring the sex-linked segment of the kiwifruit genome, the researchers have determined that the Y-encoded sex-determinant genes *Shy Girl* (*SyG1*) and *Friendly Boy* (*FrBy*) act independently as the suppressors of feminization and promoters of male factors, respectively. Non-expression of *SyG1* in males results in the formation of hermaphrodites; yet, masculine behaviour has been reported to develop in this fruit crop with plunking of *FrBy* gene into the female plants. These findings have created new avenues for generating horticulturally important self-pollinating hermaphrodite vines, eliminating the need to waste orchard space on non-fruiting male varieties.

*Corresponding author: E-mail: shashi9uhf@gmail.com, shashi_uhf@yahoo.com;



Keywords: Dioecy; inconstant male; (A) BCE floral model; shy girl; friendly boy.

1. INTRODUCTION

Kiwifruit (*Actinidia deliciosa*) is a rapidly developing fruit crop from one side of the planet to the other. Currently, it is grown across more than 270457 hectares worldwide [1]. In 2021, the global production of kiwifruit reached 4.47 million tonnes, with China, New Zealand, Italy, Greece, Iran and Chile leading the production [2]. Over the previous decade, its exports has seen an average increase 20.1% among all exporting nations. Inferable from its nutritional importance and demand, the market share of this fruit is

expected to increase to USD 2.42 billion by 2029, at a CAGR of 5.02% [3]. Moreover, its inherent taste, nutritional content, and storage capabilities are contributing to its escalating popularity. The increasing awareness among consumers regarding its health advantages is persuading a broader population globally to regularly consume kiwifruit as part of a balanced diet. Recent figures indicate that as kiwifruit consumption grows in regions such as Asia, the European Union, and the United States, its demand is projected to remain strong for several decades [4].

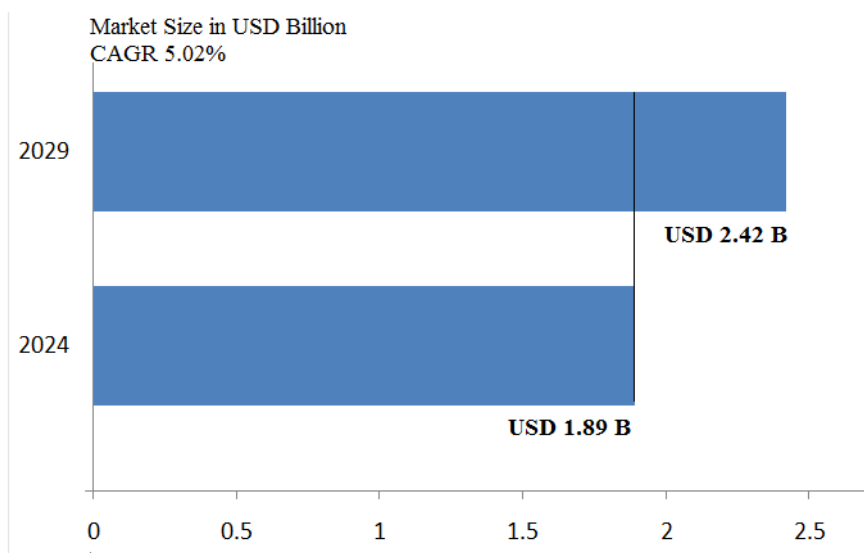


Fig. 1. Kiwifruit market
Source: Mordor Intelligence [3]

The demand-driven production goals for kiwifruits are pushing global researchers to develop varieties that can offer higher returns per unit area of cultivation. Present-day growers are seeking types that can self-pollinate and eliminate the need for specific type pollinators and pollinizers. Thus, self-pollinating varieties are anticipated to simplify orchard management and optimize the use of land in kiwifruit vineyards. Even though there are few self-pollinating varieties (inconstant male varieties or female varieties with lower proportion of viable pollen), they too produce more reliable crops when pollinated by the pollen from the male kiwifruit plants [5,6,7]. Therefore, building knowledge in kiwifruit flowering and the floral organs development is projected to represent a significant advancement towards creating hermaphroditic genotypes.

Hermaphroditism is a condition in which both male and female sex organs are present on each flower born on a plant of a particular species. It is the most common sexual state, and only a few species (5 to 6%) have evolved to bear sex-explicit plants (dioecious plants) [8]. Asparagus, papaya, persimmon, cannabis, and kiwifruit are examples of dioecious flowering plants. Most of these species are distantly related, reflecting that these plants digressed from hermaphroditism several times throughout their transformative history. Most flowering plants are otherwise hermaphroditic or monoecious, with both male and female flowers occurring on the same plant; although androdioecy, gynodioecy or trioecy are also not uncommon. But in kiwifruit, dioecy is quite dominant. According to the literature, scientists around the world are making great progress in understanding kiwifruit flowering to elucidate the sexual differentiation process of this fruit crop and these efforts are providing a great impetus to promote the development of hermaphrodite varieties of *Actinidia* species. This mini-review therefore aims to highlight kiwifruit hermaphroditism, which is otherwise less known among many working groups of this fruit crop.

2. FLORAL BIOLOGY AND CYTOGENETICS OF *Actinidia*

The most notable species in kiwifruit are *Actinidia deliciosa* and *Actinidia chinensis*, which produce kiwifruits, *Actinidia arguta* (kiwi berry), *Actinidia polygama* (silver vine), and the ornamental *Actinidia kolomikta* [9,10,11]. Studies have revealed that *A. chinensis* and *A. deliciosa* are diploid and hexaploid, respectively [12].

However, Ferguson [13] revealed that these two species are recognized as two assortments of the same species, *A. chinensis* var. *chinensis* and *A. chinensis* var. *deliciosa*, due to the presence of clines and extensive introgressive hybridization at places where they both grow. The best-known kiwifruits produced today are *Actinidia chinensis* var. *Deliciosa*, also known as *Actinidia deliciosa*. This species developed during the twentieth century, and in the last fifty years it has attained the status of an economically important crop across the globe.

Paleobotanical studies indicate that the genus *Actinidia* is approximately 20-26 million years of age [14]. All individuals from this genus (enveloping at least 76 species) are practically dioecious with male and female flowers being born on separate plants [15,16]. The male vines bear apparently staminate flowers with viable pollen and a rudimentary ovary that lacks style and does not form ovules. On the other hand, pistillate flowers with functionally well developed ovaries containing numerous ovules are borne on the female vines. These flowers apparently bear fully- developed stamens, which, however, produce nonviable pollen due to impaired intine wall synthesis [17].

All the *Actinidia* species possess a basic chromosome number of $x=29$, although diploid, tetraploid, hexaploid and even octaploid species are also not uncommon [18,16]. McNeillage [19,20]. Reported that a single determinant controlling sex at any ploidy level is the presence of gender inconstant males (that is, males producing occasional fruits), as the sex ratios they obtained after crossing males to females were invariably 1:1 and 3:1 when selfed. The male sex in this fruit crop is thus reported to be heterozygous (XY), and the female sex must be homozygous (XX) [21,22,23]. However, necessarily, this does not signify the presence of differential sex chromosomes, as was observed in some dioecious species by Charlesworth [24]. Furthermore, a single genetic determinant always controls sex with a unique set of X/Y chromosomes. The probable presence of a sex-neutral (XX) n chromosome set in *Actinidia* has been reported, where n depends on ploidy level [25,22,26,27].

3. SEX DIFFERENTIATION

The identity of floral organs in this genus is also determined mainly by the popular (A)BCE-like floral model [28,29,30], where the A function is

not clear [31], and the B- and C-functions act after the establishment of meristem identity to determine organ identity. Only the expression domains of the B- and C- functions align with an organ identity function in two adjacent whorls and have both individual and combined roles in establishing floral organ identity exclusively [30]. Caporali et al. [32] identified 7 distinct stages of floral development in kiwifruit. They stated that the sex differences in kiwi flower development become evident from the initial stage (stage 3) of floral development around the 10th day from the initiation of stamen development when flowers are about 3.5 to 4.5 mm in length. It was reported to be distinguishable as a compacted gynoecium in female flowers; the staminate flowers lacked the carpel meristem and ovule primordium. They further speculated that pollen degeneration in female flowers occurs due to programmed cell death at later stages, although pollen release in staminate flowers was reported to occur after that during the 7th stage of floral development. As these 7 developmental stages were linked to specific morphological differences, this provided the required information for gene expression studies related to sex differentiation and other floral developmental processes in this fruit plant.

Previously, McNeilage [20] described the characteristics of fruiting males along with those of female and pure male genotypes based on variations in numerical and morphometric floral characteristics. He reported that fruiting males produced both staminate and bisexual flowers in both uniform and mixed inflorescences. The proportion of hermaphrodite on such vines reportedly varies from 40 to 70%. It was reported that all the floral organs do not develop similarly in all the three (pistillate, staminate and hermaphrodite) sex forms; differences were reported in stylar length, ovary length, ovary diameter and the number of ovules per carpel. Staminate and bisexual flowers were reported to have the same number of flowers per inflorescence, length of filaments, pollen viability, and length of the rachis, carpel number but differed from the female flowers. Regardless of sex, all the flowers had similar number of sepals and petals. Furthermore, McNeilage and Steinhagen [33], in a rare progeny of crosses in between Hayward and an inconstant (fruiting) male, obtained complete hermaphroditic flowers with no restrictions on selfing; however, fruits were not of commercial importance.

4. FRIENDLY BOY AND SHY GIRL GENES

Flowering plants exhibit a normal tendency toward hermaphroditism, and separate male and female sex (dioecious) plants are rare. However, recent findings suggest otherwise, stating that, evolutionary dioecy is reversible when species move in and out of dioecy at a certain frequency [34]. Theories explaining the shift between hermaphroditism and dioecy predict the occurrence of at least two mutations for evolving separate sexes – the one that is responsible for male sterility and the other for female sterility. The independent evolution of dioecy in several species indicates that each transition to dioecy may involve different genes and evolutionary scenarios [35]. Therefore, plants with distinct male and female sexes are currently thought to be primitive in the genus *Actinidia*. After exploring the sex-linked segment of the kiwifruit genome Akagi et al. [36,37] reported that dioecy in *Actinidia* appeared 20 million years ago. After analyzing the transcripts from young flowers, they explained that a Y-encoded sex determinant candidate gene acting as the suppressor of feminization (SuF) is a male-specific type-C cytokinin response regulator that is expressed in developing male flowers [38]. These authors named this response regulator gene *Shy Girl* (*SyGI*). This gene gets expressed at the surface of the rudimentary carpel in male flowers and suppresses carpel development. However, the gene responsible for pollen sterility in female flowers could not be identified during these studies. Later, Akagi et al. [36] identified a second Y-encoded sex determinant, named *Friendly Boy* (*FrBy*), which was reported to act to maintain male functions independently of *SyGI*. Genome re-sequencing of natural hermaphrodite flowers revealed that a hermaphrodite flower is genetically male, but partial deletions in the Y chromosome, including those of *SyGI*, have led to hermaphroditism. Furthermore, it was found that the incorporation of *FrBy* also resulted in hermaphroditism; when a copy of *FrBy* was plunked into the genome of a female kiwifruit plant; within months the plants produced hermaphrodite flowers [36]. These findings clearly indicated that Y-encoded *SyGI* and *FrBy* act independently as the suppressor of feminization (SuF) and promoter of male factors (M), respectively, confirming the presence of a two-factor sex determination system for the evolution of gynodioecy from hermaphrodite or monoecious individuals, as explained earlier by Charlesworth and Charlesworth [39]. This elucidation of the mechanisms underlying sex

determination has extended the possibilities of manipulating the *SyG1* and *FrBy* genes to generate self-pollinating hermaphrodite cultivars in Kiwifruit.

5. HORTICULTURAL SIGNIFICANCE OF HERMAPHRODITISM

Hermaphrodite plants may benefit from reproductive assurance, efficient pollinator attraction and the ability to optimize resource allocation to boost regenerative accomplishment during the blooming period. Hermaphroditism dodges the requirement for a mating partner. In addition, the transition from dioecy to hermaphroditism in kiwifruit is supposed to dispense with the need to plant non-fruiting pollinizer cultivars. When pollinator activity is curtailed due to harsh weather, pollination and fruit set are typically significant concerns. However, planting bisexual cultivars can significantly reduce pollinator-dependent fruitset issues. Furthermore, through hermaphrodite vines, efficient pollination and fertilisation by selfing typically yield higher seed numbers and, consequently, superior fruit size and quality [40,41,42,43].

6. CONCLUSION

Since hermaphroditism is one of the most desired traits in both monoecious and dioecious fruit crops, researchers have always been attracted to it. *Actinidia deliciosa*, or kiwifruit, is not always strictly dioecious; males planted in vineyards as pollinizers sometimes produce both staminate and bisexual flowers and produce fruits of poor quality. In order to develop commercially important hermaphroditic vines, the scientists now have decoded the process of kiwifruit sex differentiation which is determined by the independent expression of *SyG1* and/or *FrBy* genes. In male plants development of carpel is prevented by the expression of *SyG1*. The degeneration of pollens on the other hand occurs due to programmed cell death in female vines. However, plunking of *FrBy* in such plants has been reported to induce hermaphroditism. It has therefore been speculated that the manipulation of *SyG1* and/or *FrBy* can potentially change the dioecy of elite cultivars of kiwifruit to hermaphroditism. This would provide kiwifruit vines with high reproductive assurance, eliminating the need for pollinizers or pollinators. In the future, *orchard productivity could be increased by planting only elite hermaphrodite*

vines, negating the need to plant any male vines that *are fruitless*.

COMPETING INTERESTS

Author has declared that no competing interests exist.

REFERENCES

1. UNDATA; 2020. Available:<http://data.un.org/Data.aspx?d=FAO&f=itemCode%3A592> (accessed on 27 November, 2023).
2. Shahbandeh M. Global kiwi production 2000-2021. Statista; 2023. Available:<https://www.statista.com/statistics/577896/world-kiwi-production/>
3. Mordor Intelligence. Kiwi fruit market size & share analysis - Growth trends & forecasts (2024 - 2029). Mordor Intelligence; 2023. Available:<https://www.mordorintelligence.com/industry-reports/kiwi-fruit-market>
4. Eurofruit. Kiwifruit demand 'to remain positive'; 2022. Available:<https://www.fruitnet.com/eurofruit/kiwifruit-demand-to-remain-positive/187979.article>
5. Bellini E, Rotundo A, Pilone N. Observations on self-fertile clones of kiwifruit. New Zealand Journal of Crop and Horticultural Science. 1991;19:337-339.
6. Morton J. Kiwifruit: *Actinidia deliciosa*. In: Fruits of warm climates, 1987. Centre for new crops & plant products at purdue university; 2011. Retrieved 17 January, 2024. Available:http://www.hort.purdue.edu/newcrop/morton/kiwifruit_ars.html
7. Pinto T, Vilela A. Kiwifruit, a botany, chemical and sensory approach – A review. Advances in Plants and Agriculture Research. 2018;6:383-390.
8. Casimiro-Soriguer I, Buide ML, Narbona E. Diversity of sexual systems within different lineages of the genus *Silene*. AoB Plants. 2015;7:plv037. DOI: 10.1093/aobpla/plv037 PMID: 25862920 PMID: PMC4433491
9. Ferguson AR. New temperate fruits: *Actinidia chinensis* and *Actinidia deliciosa*. Perspectives on New Crops and New Uses. 1999;342-347.
10. Campbell-Culver M. The origin of plants. The people and plants that have shaped

- Britain's garden history since the year 1000. London: Headline. 2001;69. 22.
11. Marner S. Plant 304, *Actinidia* species (Actinidiaceae). *Kiwifruits*. Oxford Plants 400; 2023.
Available:<https://herbaria.plants.ox.ac.uk/bol/plants400/Profiles/ab/Actinidia> (accessed on 27 November, 2023).
 12. Liang C-F, Ferguson AR. Emendation of the Latin name of *Actinidia chinensis* Pl. vat. hispida C. F. Liang. *Guihaia*. 1984;l: 181-182.
 13. Ferguson AR. Botanical description. In: Testolin, R., Huang, H.-W. and Ferguson, A.R. (eds) *The Kiwifruit Genome*. Springer, Berlin. 2016;1–13.
 14. Qian YQ, Yu DP. Advances in actinidia research in China. *Acta Horticulturae*. 1991;297:51-55. ISSN 0567-7572.
 15. Ferguson AR. Kiwifruit (*Actinidia*). *Acta Horticulturae*. 1990;290:601-653. ISSN 0567-7572.
 16. Ferguson AR, Huang HW. Genetic resources of kiwifruit: domestication and breeding. *Horticultural Reviews*. 2007;33: 1–121.
 17. Messina R. Microsporogenesis in male-fertile cv. Matua and male-sterile cv. Hayward of *Actinidia deliciosa* var. *deliciosa* (kiwifruit). *Advances in Horticultural Sciences*. 1993;7:77-81.
 18. McNeilage MA, Considine JA. Chromosome studies in some *Actinidia* taxa and implications for breeding. *New Zealand Journal of Botany*. 1989;27:71–81.
 19. McNeilage MA. Gender variation in *Actinidia deliciosa*, the kiwifruit. *Sexual Plant Reproduction*. 1991a;4:267–273.
 20. McNeilage MA. Sex expression in fruiting male vines on kiwifruit. *Sexual Plant Reproduction*. 1991b;4:274–278.
 21. Testolin R, Cipriani G, Costa G. Sex segregation ratio and gender expression in the genus *Actinidia*. *Sexual Plant Reproduction*. 1995;8:129–132.
Available:<https://doi.org/10.1007/BF00242255>
 22. Testolin R, Cipriani G, Messina R. Sex control in *Actinidia* is monofactorial and remains so in polyploids. In: Ainsworth, C.C. (ed.) *Sex Determination in Plants*. Bios Scientific Publishers. Oxford. 1999; 173–181.
 23. Harvey CF, Gill GP, Fraser LG, McNeilage MA. Sex determination in *Actinidia*. 1. Sexlinked markers and progeny sex ratio in diploid *A. chinensis*. *Sexual Plant Reproduction*. 1997;10: 149-154.
 24. Charlesworth D. Plant sex determination and sex chromosomes. *Heredity*. 2002; 88:94-101.
 25. McNeilage MA. Progress in breeding hermaphrodite kiwifruit cultivars and understanding the genetics of sex determination. *Acta Horticulturae*. 1997; 444:73-78.
Available:<https://doi.org/10.17660/ActaHortic.1997.444.8>
 26. Testolin R, Messina R, Lain O, Cipriani G. A natural sex mutant in kiwifruit (*Actinidia deliciosa*). *New Zealand Journal of Crop and Horticultural Science*. 2004;32(2):179-183.
 27. Fraser LG, Tsang GK, Datson PM, De Silva HN, Harvey CF, Gill GP, Crowhurst RN, McNeilage MA. A gene-rich linkage map in the dioecious species *Actinidia chinensis* (kiwifruit) reveals putative X/Y sex-determining chromosomes. *BMC Genomics*. 2009;10(1):1-15.
 28. Coen ES, Meyerowitz EM. The war of the whorls: Genetic interactions controlling flower development. *Nature*. 1991;353 (6339):31-37.
 29. Bowman JL. Evolutionary conservation of angiosperm flower development at the molecular and genetic levels. *Journal of Biosciences*. 1997;22:515-527.
 30. Causier B, Schwarz-Sommer Z, Davies B. February. Floral organ identity: 20 years of ABCs. In *Seminars in cell & developmental biology*. 2010;21(1):73-79. Academic Press.
 31. Varkonyi-Gasic E, Moss SM, Voogd C, Wu R, Lough RH, Wang YY, Hellens RP. Identification and characterization of flowering genes in kiwifruit: Sequence conservation and role in kiwifruit flower development. *BMC Plant Biology*. 2011; 11:1-16.
 32. Caporali E, Testolin R, Pierce S, Spada A. Sex change in kiwifruit (*Actinidia chinensis* Planch.): a developmental framework for the bisexual to unisexual floral transition. *Plant Reproduction*. 2019;32:323-330.
 33. McNeilage MA, Steinhagen S. Flower and fruit characters in a kiwifruit hermaphrodite. *Euphytica*. 1998;101(1):69-72.
 34. Käfer J, Marais GAB, Pannell JR. On the rarity of dioecy in flowering plants. *Mol. Ecol*. 2017;26:1225–41.
 35. Renner SS. The relative and absolute frequencies of angiosperm sexual

- systems: Dioecy, monoecy, gynodioecy, and an updated online database. *Am. J. Bot.* 2014;101(10):1588–1596.
36. Akagi T, Pilkington SM, Varkonyi-Gasic E, Henry IM, Sugano SS, Sonoda M, Firl A, McNeilage MA, Douglas MJ, Wang T, Rebstock R. Two Y-chromosome-encoded genes determine sex in kiwifruit. *Nature Plants.* 2019;5(8):801-809.
37. Akagi T, Henry IM, Ohtani H, Morimoto T, Beppu K, Kataoka I, Tao R. A Y-encoded suppressor of feminization arose via lineage-specific duplication of a cytokinin response regulator in kiwifruit. *The Plant Cell.* 2018;30(4):780-795.
38. Varkonyi-Gasic E, Wang T, Cooney J, Jeon S, Voogd C, Douglas MJ, Pilkington SM, Akagi T, Allan AC. Shy girl, a kiwifruit suppressor of feminization, restricts gynoecium development via regulation of cytokinin metabolism and signaling. *New Phytologist.* 2021;230(4):1461-1475.
39. Charlesworth B, Charlesworth D. A model for the evolution of dioecy and gynodioecy. *American Naturalist.* 1978;112:975–997.
40. Awasthi RP, Kumar Shashi. Kiwifruit. In: Verma, I. R. And Jindal, K. K. (eds.) fruit crops pollination, Kalyani Publishers, Ludhiana, India. 1997;227-237.
41. Gianni T, Vania M. Artificial pollination in kiwifruit and olive trees. *Pollination in Plants. InTech*; 2018. Available:<http://dx.doi.org/10.5772/intechopen.74831>
42. Castro H, Siopa C, Casais V, Castro M, Loureiro J, Gaspar H, Castro S. Pollination as a key management tool in crop production: Kiwifruit orchards as a study case. *Scientia Horticulturae.* 2021;290:110533. Available:<https://doi.org/10.1016/j.scienta.2021.110533>
43. McNeilage MA. Gender variation in *Actinidia deliciosa*, the kiwifruit. *Sexual Plant Reproduction.* 1991;4:267-273.

© Copyright (2024): Author(s). The licensee is the journal publisher. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<https://www.sdiarticle5.com/review-history/114445>